OXALIS Diagnostics Pvt. Ltd.

Technical Data

CLED Agar with Bromothymol Blue. 100 g / 500 g
Agar with Bromothymol Blue is recommended for isolation, enumeration and presumptive identification of urinary pathogens on the basis of lactose fermentation.

Product Presentation:

Cat No.	Product description	Pack Size
11030030100	CLED Agar with Bromothymol Blue	100 Gram
11030030500	CLED Agar with Bromothymol Blue	500 Gram

Principle
L-Cystine lactose electrolyte deficient medium composed of peptone, tryptone, meat extract, lactose, L-Cystine and bromothymol blue. Literature suggests that the *Proteous* species can be controlled by restricting the electrolytes and by replacing the mannitol by lactose and sucrose with L-Cystine and bromothymol blue. Peptone, meat extract and tryptone serve as the source of all essential nutrients such as amino acids, vitamins, other trace factors. L-Cystine is added as a growth supplement for cystine-dependent coliforms. Lactose is included as a carbon source and plays a crucial role for selection of lactose fermenting microbes. Brom Thymol Blue is used as a pH indicator. Organisms capable of fermenting lactose will lower the pH of medium, result in change the color of the medium from green to yellow. Agar is used as a solidifying agent.

Composition

Ingredients	Grams / Litre
Meat Extract	3.00
Peptone	4.00
Tryptone	4.00
Lactose	10.0
L-Cystine	0.128
Bromothymol Blue	0.02
Agar	15.0

Final pH (at 25°C) 7.3± 0.2
*Formula adjusted, standardized to suit performance parameters

Type of specimen
Clinical samples - Urine

Specimen Collection and Handling
Ensure that all samples are properly labeled. Follow appropriate techniques for handling samples as per established guidelines. Some samples may require special handling, such as immediate refrigeration or protection from light, follow the standard procedure. The samples must be stored and tested within the permissible time duration. After use, contaminated materials must be sterilized by autoclaving before discarding.

FACTORY & OFFICE
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Directions

- ✓ Suspend 36.00 g of powder in 1000 mL distilled water.
- ✓ Mix thoroughly.
- ✓ Boil to dissolve the medium completely. Avoid Overheating.
- ✓ Sterilize by autoclaving 121°C for 15 minutes or as per validated cycle.

Storage and Stability

- ✓ Store Dehydrated culture media in cool, dry place at 10°C-30°C away from direct light.
- ✓ Store prepared medium at 2°C-8°C. Avoid freezing and overheating. Use before expiry date on the label. Once opened keep powdered medium closed to avoid hydration.

Quality Control

Dehydrated Appearance: Beige with slight green tint coloured, homogenous, free flowing powder.
Prepared Appearance: Green to blue green coloured, clear to slightly opalescent gel forms in petridishes.
Growth Promotion Test: Cultural characteristics observed after an incubation of 18-48 hours at 35°C -37°C.
Cultural Response :

Organism	Type Culture	Growth	Colour of the Colany	Incubation Temperature	Incubation Period
Escherichia coli	ATCC 8739	Good	Yellow	35°C -37°C.	18-48 Hours
Staphylococcus aureus	ATCC 25923	Good	Yellow	35°C -37°C.	18-48 Hours
Salmonella Typhi	ATCC 14028	Good	Blue	35°C -37°C.	18-48 Hours

Interpretation of Results

- ✓ Count the number of colonies on the dipstick. Multiply by dilution factor to convert the count to CFU per mL of the sample.
- ✓ Contaminant bacteria usually appear in low numbers, which vary in colony morphology.

Warranty

- ✓ This product is designed to perform as described on the label and package insert. The manufacturer disclaims any implied warranty of use and sale for any other purpose.

Disposal

Disposal of infectious material and material that comes in to contact with clinical sample must be decontaminated and dispose of by autoclaving or incineration or established laboratory procedures.
User must be ensure safe disposal of used or unusable preparation of the products.

Reference

1. Mackey and Sandys, 1965, Br. Med. J., 2:1286.
2. MacKey and Sandys, 1966, Br. Med. J., 1:1173.
3. Dixson J. M. S. and Clark M. A., 1968, Conc. Med. Assoc. J., 99 (15)
4. Benner E. J., 1970, Appl. Microbiol., 19(3), 409
5. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.

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